

In Vitro Antibacterial Activity of *Thymus satureioides*, *Mentha pulegium*, and *Origanum vulgare* Essential Oils Against *Escherichia coli* Isolated from Raw Sheep Milk

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Abstract – This study aims to evaluate *in vitro* the antibacterial activity of essential oils extracted from *Thymus satureioides*, *Mentha pulegium* and *Origanum vulgare* against *Escherichia coli* isolated from raw sheep milk of Sardi breed using dilution technique in liquid medium in order to determine MIC and MBC and compared to the effectiveness of those extrats on *E. coli* isolated from patients and sheep meat.

The yields of essential oils obtained by steam distillation were determined relatively to the plant dry matter and each oil was analyzed by gas chromatography coupled to mass spectrometry (GC-MS) in order to determine its chemical composition.

Results obtained showed that the most important yield was obtained with *M. pulegium* (3.2%) followed by *T. satureioides* (1.85%) and *O. vulgare* (1,6%).

Essential oils tested proved a very significant antibacterial activity against *Escherichia coli* strains. The great effect was observed with *T. satureioides* followed by *O. vulgare* and *M. pulegium* from low concentrations. The determination of the Minimal Bactericidal Concentrations (MBCs) has shown that plants' extracts studied have a bactericidal effect on all strains and their values in mg/ml are identical to those of the Minimal Inhibitory Concentrations (MICs).

The results of the essential oils' chemical compositions demonstrated that components of these oils are monoterpene oxygenated derivatives, monoterpene hydrocarbons and phenols.

The great antibacterial activity, proved by essential oils tested during this research, opens up an area for the use of medicinal plants as preservatives in dairy industry and for therapeutic purposes.

Keywords – Sheep Milk, Antibiotic Resistance, Essential Oils, Medicinal Plants, *Escherichia coli*.

I. INTRODUCTION

Food security, in which the microbiological quality of food is an essential component, represents a considerable challenge. Diseases caused by collective food poisoning (TIAC) represent globally a significant number of deaths in developing countries.

In Europe, mortality due to food poisoning is low, but a number of 50000 acute gastroenteritis per million inhabitants per year is commonly cited [1]. In Morocco, the majority of cases of food poisoning have been reported more than 86% of bacterial origin including meat [2] and milk which are the most affected foods [3] – [4].

Antibiotics are among the most successful drugs used to cure human and animal infections caused by pathogenic bacteria [5] – [6]. Unfortunately, the hopes raised by the discovery of those pharmaceutical products were quickly reconsidered given the ability of bacteria to develop resistance even of optimal use [7]. The prevalence of antibiotic resistance among food borne pathogens has then increased in recent decades [8] because of the increase in the number of antibiotic-resistant Gram-negative bacteria especially *Escherichia coli* [9] – [10] – [11] – [12] – [13].

Recent studies [14] – [8] – [5] have demonstrated that the patterns of antibiotic usage may greatly affect the number of resistant microorganisms which occurs in an environment. An earlier study in Kenya [15] found betalactam antibiotics residues to be prevalent in milk. Evidence has been found which indicates that resistance strains of pathogens can be transmitted to humans through food [16] – [17].

On the other hand, several questions are currently raised about the safety of chemicals used in medicine or food

industry. Many substances produced during the manufacturing process and storage of food under the action of free radicals lead to oxygenated change of taste, odor and color and consequently loss of quality and food safety.

The human use of aromatic and medicinal plants and essential oils is so ancient. They are used traditionally to ward off bad luck, to heal, for relaxation, food flavoring and preserving of food [18]. Therefore, many research groups were interested in the functional role plant extracts especially essential oils play. These latter are appreciated for their bioactive efficacy as fungicides [19], bactericides [20], anti-inflammatory [21], antiviral [22] and antioxidants [23].

These last years, the use of ethnobotanical information in the search for medicinal plants has greatly attracted attention from the scientific community [24]. The search for biologically active extracts from plants traditionally used is relevant and has been highlighted by Leaman *et al.* (1995) [25] in their study of plants traditionally used in Borneo in the treatment of malaria. This approach has also led to the discovery of rich sources of compounds with antiviral and antibiotic activities [26] – [27] and then molecules which can be used as food preservatives [28].

In developing countries, such as Morocco and rural societies, the use of medicinal plants is both a necessity and a valuable resource and also provides a real alternative for systems of care and health. Thanks to its special geographical position, Morocco enjoys a bioclimate for a rich and diverse vegetation. More than 4200 species and subspecies have been identified, including a hundred endemic and more than 9% of the total Moroccan flora are used as medicinal plants and aromatic interests.

Our study is part of a plant resource development in Morocco and the improvement of the hygienic quality of milk in order to identify new food preservatives available locally, against pathogens commonly involved in food contamination. It aims to assess the *in vitro* resistance or susceptibility of 10 *Escherichia coli* strains isolated and collected manually from raw sheep milk of Sardi breed in Morocco to *Thymus satureioides*, *Mentha pulegium* and *Origanum vulgare* essential oils; they were compared to 5 *Escherichia coli* strains isolated from patients and 4 strains from raw sheep meat.

II. MATERIAL AND METHODS

1) Plant material

Harvest of *Thymus satureioides*, *Mentha pulegium* and *Origanum vulgare* samples was made between May and July 2011: *T. satureioides* was harvested in the region of Agadir; *vulgare* in Masmoda and *M. pulegium*, in Ouazzane (Central Morocco). These species were identified by Dr. Aafi, a botanist at Forest Research Centre of Rabat, Morocco.

2) Extraction of essential oils

The extraction of essential oils from aerial part (stems, leaves and flowers) was performed by steam distillation using a Clevenger-type apparatus [29]. Three distillations were carried out by boiling, for one hour thirty minutes;

200 g of fresh plant material was placed in 1 L water inside a 2 liter flask. The essential oil yield was determined in relation to the dry matter and estimated from three dried samples (30 g) for 48 h in an oven at 60°C. The essential oil was then stored at 4°C in the dark in the presence of anhydrous sodium sulfate [30].

3) Microorganisms tested

10 *Escherichia coli* strains were isolated from the raw sheep milk on Desoxycholate Lactose Agar (DL, Oxoid, England) culture medium (selective culture medium to detect bacteria belonging to the family Enterobacteriaceae). The identification of *E. coli* strains was performed by determining the biochemical profile using respectively the kit API 20 E (Biomerieux, France). Seeding and reading these kits were made according to the manufacturer's instructions. The identification of each isolate was obtained using the Api Plus software [31] – [32] – [33] – [34] – [35].

Escherichia coli 1, *Escherichia coli* ATCCS, *Escherichia coli* 2, *Enterohemorrhagic Escherichia coli* (EHEC) O157 and *Enteropathogenic Escherichia coli* (EP) isolated from humans and *Enterotoxinogenic Escherichia coli* (ETEC), *Enteropathogenic Escherichia coli* (EPEC), *Enterogregatif Escherichia coli* (EAggrec) and *Enteroinvasive Escherichia coli* isolated from raw sheep meat; in the Bacteriology Laboratory of the National Institute of Hygiene of Rabat were also tested.

4) Evaluation of the antibacterial activity of essential oils

With the evaluation of antimicrobial activity, we aim to determine the antibacterial parameters (minimum inhibitory concentration (MIC) and minimum bactericidal concentration (MBC)) of different essential oils studied. To do this, we used the method of broth microdilution coupled with the spread on solid medium as described by Chabbert and Daguet [36].

Determination of MIC essential oils against bacterial strains was done by microtiter technique with flat bottom sterile plates (Bio-Rad), as described by Eloff (1998) [37]; ; tetrazolium (MTT: 3 - (4,5 - dimethylthiazol-2-yl) - 2,5 diphenyltetrazolium bromide (Sigma, St. Louis, MO) was used as an indicator of viability.

100 µl of BHI (Brain Heart Infusion) was deposited at each well, and 90 µl of the essential oil mixed with Tween 80 or successive dilution of 1/2 was then added to each well. The latter was then inoculated with 10 µl of a microbial suspension (equivalent to 10⁶ cells / ml).

After incubation at 37°C for 24 h, 10 µl of the MTT, freshly prepared with 0.4 mg / ml of sterile saline, was added to each well. The plate was incubated again for 10 to 30 min at 37°C. Wells, where growth occurred, showed a blue-violet color. Negative controls were prepared in isolated wells by adding strains tested in only the adequate culture medium without extract.

In addition, the determination of the MBC requires streaking 100 µl of the wells contents at a concentration greater than or equal to the MIC in the dilution series previously established on nutrient agar. Thus, the MBC was determined after incubation for 24 h at 37°C. This was the lowest concentration that completely inhibited growth.

5) Chromatographic analyses

Chromatographic analysis was performed on a gas chromatography with electronic pressure control type Hewlett Packard (HP 6890 series) equipped with a capillary column HP-5 (30 m x 0.25 mm), having a film thickness of 0.25 microns, an FID detector set at 260°C and fed with a mixture of gases and an H₂/Air split-splitless injector set at 275°C. The injection mode was split (split ratio: 1/50). The gas used was nitrogen with a flow rate of 1.7 ml min⁻¹. The column temperature was programmed from 50 to 250°C at 4°C min⁻¹. The device is controlled by a computer system type "HP ChemStation" that manages the operation of the device and monitors the chromatographic analysis. The identification of the components was carried out on the basis of Kováts indices (IK) and the gas chromatography coupled to mass spectrometer (GC-MS). The latter is performed on a gas chromatography type Hewlett Packard (HP 6890 series) coupled to a mass spectrometer (HP 5973 series). Fragmentation is performed by electron impact at 70 eV. The column used was a HP-5MS capillary column (30 m x 0.25 mm), with film thickness of 0.25 microns. The column temperature was programmed from 50 to 250°C at 4°C min⁻¹. The carrier gas was helium with a flow rate of 1.5 ml min⁻¹. The injection mode was split mode (split

ratio 1/70). The device was connected to a computer system that manages a library of mass spectra NIST 98 [37].

III. RESULTS

1. Essential oils yields

Average yields of essential oils were calculated in milliliters relative to 100 g of dry plant material. These rates are 1.85% for *Thymus satureioides*, 3.2% for *Mentha pulegium* and 1.6% for *Origanum vulgare*. These rates indicate that the performance of *M. pulegium* is the highest. As a result, it should be interesting to consider its use subject to good performance of the antibacterial activity.

2. Antibacterial activity of essential oils tested

The results of the antibacterial effect test of different essential oils used are summarized in Figure 1. Note that all essential oils have an inhibitory activity against the bacteria tested.

Moreover, the MICs determination of these natural extracts showed varying levels of action that could be mainly due to the chemical composition of each essential oil.

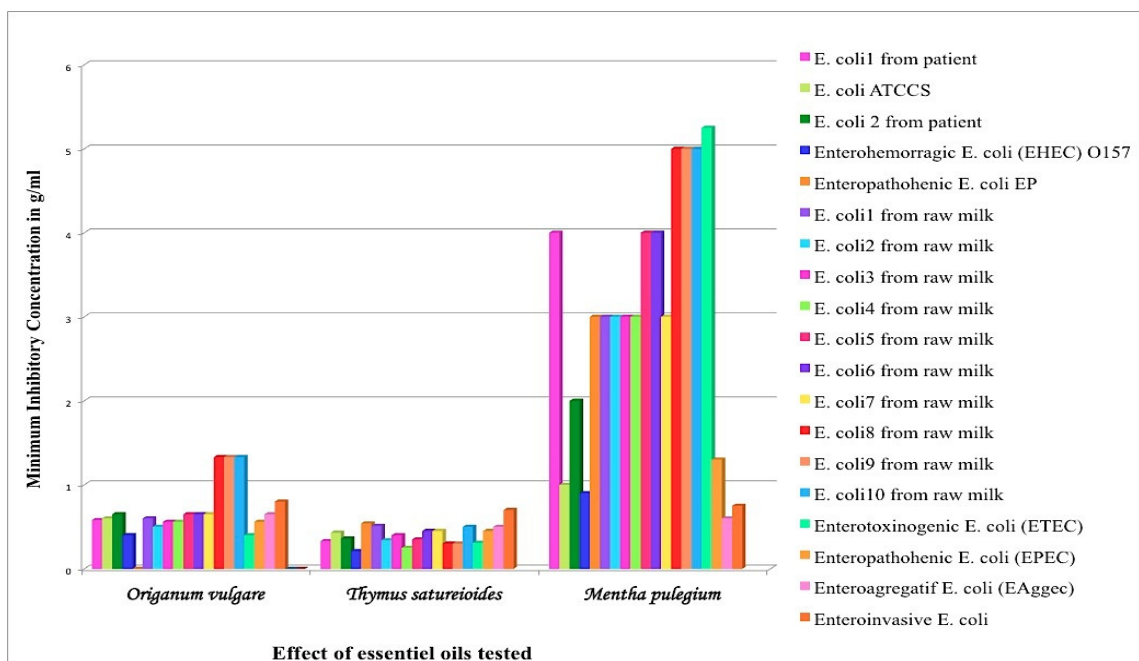


Fig.1. Minimum inhibitory concentrations of essential oils tested on the growth of *Escherichia coli* strains

The inhibitory effect of these extracts also proved to be bactericidal against all species studied (Table I). Analysis of the antibacterial test results obtained *in vitro* with the four essential oils shows a remarkable efficiency of these natural products against pathogenic strains studied with difference in efficacy depending on the species and essential oil dilutions.

In Figure 1, *Thymus satureioides* essential oil showed the greatest antibacterial effect against all strains studied. Indeed, *Escherichia coli* (EHEC) O157 was the most sensitive strain with a MIC of about 0.21 mg/ml followed

by *E. Coli* isolated from raw sheep milk whose MICs range between 0.25 and 0.51 mg/ml. By cons,

Enteroinvasive E. coli has been resistant with a MIC of about 0.7 mg/ml.

Origanum vulgare essential oil showed also a great antibacterial action against the strains studied. Thus, it was more active on *E. coli* (EHEC) O 157 isolated from patients and *E. coli* (ETEC) isolated from raw sheep meat (MIC = 0.40 mg/ml), which is the most sensitive. *E. coli* from raw sheep milk was totally inhibited from 0.50, whose *E coli* 8, *E coli* 9 and *E coli* 10 were inhibited at

high concentration of 1.33 mg/ml. Whereas, the rest bacteria were inhibited from MIC of 0.65 to 0.80 mg/ml.

The determination of MICs for *Mentha pulegium* essential oil showed antibacterial activity levels that varied based on the strains tested. The essential oil of this plant could completely inhibit bacterial growth of all *E. coli* tested with MICs that fluctuate between 0.6 and 5.25 mg / ml. In the presence of the latter extract, *E. coli* (EAggec) was the most sensitive while *E. coli* (ETEC) was more resistant. *E. coli* from raw sheep milk was completely inhibited from 3 to 5 mg/ml.

Based on the MIC, we note that essential oils used have different activities against strains of *Escherichia coli*. The

activity of *Thymus satureioides* had the best performance followed by the essential oil of *Origanum vulgare* and *Mentha pulegium*, respectively.

To characterize the antibacterial activity of these essential oils, cultures of each strain that have not started growth in the presence of the plant extract were transferred to a neutral culture medium without extract. After the incubation time, we note that the antibacterial activity of all the plants' extracts is bactericidal against all strains of *Escherichia coli* studied. Indeed, no resumption of growth was observed, which proves the bactericidal effect of these extracts (Table I).

Table I : Minimum bactericidal concentration of essential oils tested on the growth of *E. coli* strains Minimum bactericidal concentrations in mg /ml

Bacteria tested	<i>O. vulgare</i>		<i>T. Satureioides</i>		<i>M. pulegium</i>	
<i>E. coli</i> 1	0.58	B	0.33	B	4	B
<i>E. coli</i> ATCCS	0.6	B	0.43	B	1	B
<i>E. coli</i> 2	0.65	B	0.36	B	2	B
<i>E. coli</i> (EHEC) O157	0.4	B	0.21	B	0.9	B
<i>E. coli</i> EP	0.6	B	0.54	B	3	B
<i>Escherichia coli</i> 1	0.5	B	0.51	B	3	B
<i>Escherichia coli</i> 2	0.5	B	0.34	B	3	B
<i>Escherichia coli</i> 3	0.56	B	0.4	B	3	B
<i>Escherichia coli</i> 4	0.56	B	0.25	B	3	B
<i>Escherichia coli</i> 5	0.56	B	0.35	B	4	B
<i>Escherichia coli</i> 6	0.65	B	0.45	B	4	B
<i>Escherichia coli</i> 7	0.65	B	0.45	B	3	B
<i>Escherichia coli</i> 8	1.33	B	0.3	B	5	B
<i>Escherichia coli</i> 9	1.33	B	0.3	B	5	B
<i>Escherichia coli</i> 10	1.33	B	0.5	B	5	B
<i>E. coli</i> (ETEC)	0.4	B	0.31	B	5.25	B
<i>E. coli</i> (EPEC)	0.56	B	0.45	B	1.3	B
<i>E. coli</i> (EAggec)	0.65	B	0.5	B	0.6	B
Enteroinvasive <i>E. coli</i>	0.8	B	0.7	B	0.75	B

B : Bactericidal effect

3. Chemical compositions

The chromatographic analysis of plants' essential oils tested revealed the presence of volatile compounds: 24 for *T. satureioides*, 23 for *M. pulegium* and 18 for *O. vulgare*, respectively. These components represent : 99.55 %,

99.7% and 97.64% of each essential oil cited, respectively. The combination of volatile compounds of these species varies in terms of diversity and concentration (Table II).

Table II: Chemical compositions of essential oils tested

N°	Ik	Component	% of components		
			<i>Thymus satureioides</i>	<i>Origanum vulgare</i>	<i>Mentha pulegium</i>
1	919	Tricyclene	0.43	-	-
2	923	α -thujene	1.18	-	-
3	930	α -pinène	6.04	0.9	0.39
4	945	Camphene	10.32	9.7	
5	952	Cyclohexanone-3-méthyl	-	-	0.28
6	959	1-octen-3-ol	-	0.9	-
7	970	Sabinene	0.20	0.5	0.45
8	973	β -pinène	1.48	0.4	0.16
9	988	Myrcène	1.14	1.4	0.99

10	1001	δ -2-carène	-	-	0.16
11	1018	α -terpinène	1.02	0.6	-
12	1019	limonene	-	0.9	1.84
13	1022	β -phyllandrene	-	0.8	-
14	1025	o-cymène	4.30	0.5	-
15	1029	p-cymène	1.24	25.9	-
16	1034	E- β -cyméne	-	0.5	-
17	1054	Menthone	-	-	0.39
18	1057	γ -terpinene	5.78	2.8	-
19	1072	p-mentha-3,8-diène	-	-	1.23
20	1084	Terpinolène	0.23	-	-
21	1096	Linalool	2.83	1.9	-
22	1098	α -thujone	-	0.9	-
23	1160	p-cymen-8-ol	-	0.8	-
24	1164	Bornéol	25.98	-	-
25	1065	Terpen-4-ol	-	0.3	-
26	-	p-menthène-5-one	-	-	1.37
27	1173	Menthol	-	-	0.74
28	1175	terpin-1-ol	1.88	-	-
29	1189	α -terpineol	8.75	0.9	-
30	1194	dihydrocarvone	-	-	2.57
31	1195	Verbanol	0.46	-	-
32	1226	Cis carveol	0.48	-	-
33	1238	R(+)-ulègone	-	-	80.33
34	1240	Carvone	0.62	-	-
35	-	Eucarvone	-	-	3.75
36	1252	pépritone	-	-	0.97
37	1283	α -Terpin-7-al	0.89	-	-
38	1288	Thymol	0.23	-	-
39	1298	Carvacrol	17.54	31.5	-
40	1365	myrtenyl acetate	-	16.4	-
41	1410	β -caryophyllene	-	0.5	-
42	1415	Caryophellene	5.98	0.5	-
43	1419	Caryophyllène	-	-	0.95
44	1453	NI	0.55	-	-
45	1509	δ -cadinene	-	0.2	-
46	1630	γ -eudesmol	-	-	0.48
47	1649	α -eudemol	-	-	0.59

IV. DISCUSSION

The use of medicinal plants with antimicrobial properties constitutes one of the most interesting avenues to explore [38]. It is in this context that we are interested in the study of the antimicrobial activity of *Thymus saturoioides*, *Mentha pulegium* and *Origanum vulgare* extracts known for their use in traditional medicine and their diverse therapeutic (anti-inflammatory, antispasmodic, sedative, antibacterial and antioxidant) activities [39].

Essential oil yields of *T. saturoioides*, *M. pulegium*, *O. vulgare* and *A. triphylla* are : 1.85%, 3.2% and 1.6%, respectively. These yields are relatively average compared to some plants that are used industrially as a source of essential oil. Our results are comparable to those obtained by El Ouali Lalami and *al* (2013) [40], who had obtained a

yield of 1.1% for *T. saturoioides* essential oil, then to those obtained by [41] who had a yield of 2.3% for *Mentha pulegium* essential oil collected from the same region, and to Souza and *al.* (2007) [42] concerning *O. vulgare* essential oil. In fact, many factors influence the performance, content, features, physico-chemical and chemical composition of essential oils as: species, environmental conditions, technical extraction, drying, time and place of harvest, tillage practices and the age of the plant [43] – [44] – [45].

Escherichia coli strains, food contamination bacteria commonly found in the digestive tract of humans and animals were tested in this study. In fact, it is known that most of *E. coli* strains are harmless. Some, however, such as *Enterohemorrhagic E. coli* (EHEC) can cause severe foodborne illness. Transmission to humans occurs mainly through consumption of contaminated foods, such as raw

ground or undercooked meat, raw milk and raw vegetables [3] – [2] – [4].

Escherichia coli isolated from raw sheep milk of Sardi breed was determined through the essential oil susceptibility on broth microdilution coupled with the spread on solid medium method. The results showed that essential oils used have, to varying degrees, antibacterial activities and inhibitory effects on *in vitro* growth of these bacterial strains.

The essential oils of *Thymus satureioides* showed an interesting antibacterial activity at the very low concentrations used (from 0.2 to 0.7 mg/ml) especially against *E. coli* (EHEC) O157. These results confirm those obtained by El Ouali Lalami and al (2013) [40], which demonstrated the effectiveness of *T. satureioides* essential oil on the growth of *E. coli* at MICs of 1.5 mg/ml. Indeed, Amarti et al. (2009) [46] reported that the extract of different species of thyme has a significant antimicrobial activity against Gram positive and Gram negative bacteria at low concentrations.

O. vulgare essential oil has completely inhibited the growth of *E. coli* at low concentrations ranging from 0.4 to 1.33 mg / ml. Our results are similar to other researches that have proved that this plant extract have a strong antibacterial activity, especially against multi-resistant bacteria as *E. coli* [47] – [44] – [41] – [48].

Analysis of *Mentha pulegium* oil components coupled with their MICs on all strains has been an interesting screening since it allows the selection of components that will be further tested separately. Then, our results showed that essential oil from this plant has an inhibitory action against bacteria studied with an antibacterial potential remarked at relatively high concentrations (0.6 to 5.25 mg/ml) compared to the previous two extracts reported. These results are consistent with the work of several researchers who have demonstrated that *M. pulegium* has an inhibitory effect against a wide range of species of bacteria involved in food intoxications [49] – [50] – [51]. According to Hmiri et al. (2011) [41], the main biologically active constituents of *M. pulegium* are monoterpenes. Those compounds could be responsible for the observed activity [51].

The *in vitro* antibacterial activities of essential oils used may be due to a synergistic effect of some components. The major essential oils' constituents are borneol (26.28%) and carvacrol (17.24%) for *T. satureioides* while the essential oil of *M. pulegium* is characterized by the presence of pulegone as the main component with a content of 80.33%. These results are similar to most of studies already carried out in Morocco [52] – [53] – [41].

The use of medicinal plants with antimicrobial properties is then a more interesting avenues to explore. It is in this context that we are interested to study also the antibacterial activity of *O. vulgare* essential, plant known for its use in Moroccan traditional medicine, specially with milk and its derivatives. The main components of *O. vulgare* essential are carvacrol and Myrtenyl. The first one presents the highest percentage with an average rate of approximately 31.5%, followed by Myrtenyl acetate with a rate of 16.5%. The high rate of carvacrol recorded, gives

this essential oil the powerful antibiotic, antiseptic and antifungal properties [54]. We understand why this natural molecule is highly sought after in the field of pharmacy and of aromatherapy.

V. CONCLUSION

Preliminary results of antibacterial study of *Thymus satureioides*, *Mentha pulegium* and *Origanum vulgare* essential oils showed that the four essential oils tested proved to have important antibacterial activities *in vitro* against highly pathogenic bacteria responsible for food poisoning. The higher antibacterial effect was observed in the case of *T. satureioides* while the lowest was observed with *M. pulegium*. These actions may be related to the presence of major constituents and a synergistic effect of some components, including terpenes and phenols.

This is because natural products tested are most effective, cheap to produce and could be analyzed easily as preservative in the food industry, and they have no side effects on the well being of humans or animals. It is advisable to conduct a thorough analysis of the action mechanisms of these essential oil compounds and a more advanced research on the synergy of basic compounds and the combination of essential oil extracts in food products. Other studies are needed to estimate the toxicity of these plants' extracts and assess their potential *in vivo*; to seek strategies for potential application of these in the conservation of milk and its derivatives and the fight against microorganisms responsible for food poisoning.

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REFERENCES

- [1] F. Käferstein, K. Motarjemi, D. W. Bettcher " Foodborn disease control: A transnational challenge," *Emerging Infectious Diseases*, 1997, 3 (4): 503 – 510
- [2] N. Cohen, H. Ennaji, M. Hassar, H. Karib, "The Bacterial quality of red meat and offal in Casablanca (Morocco) ," *Mol.Nutr.Food Res*, 2006, 50:557-562.
- [3] A. Brisabois, V. Lafarge, A. Brouillaud, M.L. de Buyser, C. Collette, B. Garin Bastuji, M. F. Thore, " Les germes pathogènes dans le lait et les produits laitiers : situation en France et en Europe," *Rev. sci. tech. Off. int. Epiz.*, 1997, 16 (1), 452-471
- [4] M. Belomaria, A. O. T. Ahami, Y. Aboussaleh, B. Elboughali, Y. Cherrah, A. Soulaymani, "Environnemental origin of collective food intoxications in Morocco: case study of the region of gharb charda bni hssen," *antropo, issn-e 1578-2603*, vol. 14, 2007, p. 83-88.
- [5] A.L. Demain and S. Sanchez, "Microbial drug discovery: 80 years of progress," *The Journal of Antibiotics*, 2009, 62, 5–16;
- [6] A.E. Van den Bogaard, N. London, C. Driessen, E.E. Stobberingh, " Antibiotic resistance of faecal *Escherichia coli* in

- poultry, poultry farmers and poultry slaughterers," *Medicine and health from Oxford*; 2001, *Journal of Antimicrobial Chemotherapy*: Volume 47, Issue 6 Pp. 763-771.
- [7] A. S. Levin, "Multiresistant ACINE TOB ACTER infections: a role for sulbactam combinations in overcoming an emerging worldwide problem," *Clin. Microbiol. Infect.*, 2002, 8:144-153.
- [8] J. Davison, "Genetic exchange between bacteria in the environment," *Plasmid*, 1999, 42 : 73-91.
- [9] M. Bonten, E. Stobberingh, J. Philips, A. Houben, "Antibiotic resistance of *Escherichia coli* in fecal samples of healthy people in two different areas in an industrialized country," *Infection*, 1992, 20: 258- 262.
- [10] N. Woodford, J.F. Turton, D.M. Livermore "Multiresistant Gramnegative bacteria: the role of high-risk clones in the dissemination of antibiotic resistance," *FEMS Microbiology Reviews*. Volume 35, Issue 5, 1999, pages 736-755
- [11] S. P. Barrett, M. A Savage, M. P Rebec, A. Guyot, N Andrews, S. B Shrimpton, "Antibiotic sensitivity of bacteria associated with communityacquired urinary tract infection in Britain," *J. Antimicrob. Chemother.*, 2000, 44: 359-365.
- [12] Erb A, Stürmer T, Marre R, Brenner H (2007). Prevalence of antibiotic resistance in *ESCHERICHIA COLI* : overview of geographical, temporal, and methodological variations *European Journal of Clinical Microbiology & Infectious Diseases*. Volume 26, Issue 2, pp 83-90.
- [13] G. Zhanel George , Mel DeCorby, Nancy Laing, Barb Weshnowski, Ravi Vashisht , Franil Tailor, Kim A. Nichol, Aleksandra Wierzbowski, Patricia Baudry , James A. Karlowsky , Philippe Lagacé-Wiens , Andrew Walkty , Melissa McCracken, Michael R. Mulvey, Jack Johnson, "Antimicrobial-Resistant Pathogens in Intensive Care Units in Canada: Results of the Canadian National Intensive Care Unit (CAN-ICU) Study," 2005-2006 *Antimicrob. Agents Chemother.* April 2008 vol. 52 no. 4 1430-1437
- [14] W. Wolfgang, "Consequences of Antibiotic Use," in *Agriculture Science Magazine Policy BIOMEDICINE Medical*, 1998, Vol. 279 no. 5353 pp. 996-997
- [15] A. Shitandi, A. Sternesjö, "Detection of antimicrobial drug residues in kenyan milk," *Journal of Food Safety*, 2001, Volume 21, Issue 4, pages 205-214
- [16] F. M. Aarestrup, H.C Wegener, "The effects of antibiotic usage in food animals on the development of antimicrobial resistance of importance for humans in *Campylobacter* and *Escherichia coli*," *Microbes and Infection* .Volume 1, Issue 8, 1999, Pages 639-644
- [17] G.G. Khachatourians, "Agricultural use of antibiotics and the evolution and transfer of antibiotic-resistant bacteria," *CMAJ*, 1998, vol. 159 no. 9
- [18] M. Charai, M. Mosaddak, M. Faid, "Chemical composition and antimicrobial activities of two aromatic plants: *origanum majorana* and *o. compactum* benth," *Journal of Essential Oil Research* ; 1996, Volume 8, Issue 6, p 657-664
- [19] K.M. Soliman, R.I. Badaea, "Effect of oil extracted from some medicinal plants on different mycotoxigenic fungi," *Food Chem. Toxicol.*, 2002, 40, 1669-1675.
- [20] M. Ghanmi, B. Satrani, A. Aafi, M.R. Isamili, H. Houti, H. El Monfalouti, K. H. Bencheqroun, M. Aberchane, L. Harki, A. Boukir, A. Chaouch, Z. Charrouf, " Effet de la date de récolte sur le rendement, la composition chimique et la bioactivité des huiles essentielles de l'armoise blanche (*Artemisia herba-alba*) de la région de Guercif (Maroc oriental) ," *Phytothérapie*, 2010, 8, 295 – 301 :
- [21] H. Ismaili, L. Milella, S. Fkih-Tetouani, A. Ildrissi, A. Camporese, S. Sosa, G. Altinier , R. Della Loggia , R. Aquino , " In vivo topical anti-inflammatory and in vitro antioxidant activities of two extracts of *Thymus satureioides* leaves," *J Ethnopharmacol.* 2004 Mar;91(1):31-6.
- [22] P. Schnitzler, A. Neuner, S. Nolkemper, C. Zundel, H. Nowack, K. Heinz Sensch, J. Reichling, "Antiviral Activity and Mode of Action of Propolis Extracts and Selected Compounds," *Phytotherapy Research Special Issue: Flavonoids and polyphenolics*, 2010, Volume 24, Issue S1, pages S20-S28,
- [23] I. Laib, M. Barkat, "Etude des activités antioxydante et antifongique de l'huile essentielle extraite des fleurs sèches de *Lavandula officinalis*," *Mémoire de Magister, INATAA, Université de Constantine*, 2011, pp. 21-69.
- [24] M. Heinrich, "Ethnobotany and its role in drug development," *Phytotherapy Research*, 2000, 14,479-488.
- [25] D.L. Leaman, J.T. Arnason, R. Yusef, H. Sangat-Roemantyo, H. Soedjito, C.K. Angerhoffer, J.M. Pezzeto, "Malaria remedies of Kenya of Apolcayan, Indonesian Borneo," *Journal of Ethnopharmacology*, 1995, 73,175-183.
- [26] D.S. Arathy, G. Vanpee; G. Belot; V. Mathew; C . DeAllie; R Sharma, "Antimicrobial drug resistance in *escherichia coli* isolated from commercial chicken eggs in Grenada, West Indies," *West Indian med. j.* 2011 vol.60 no.1 Mona Jan.
- [27] J.B. Hudson "The phytochemical approach to antiviral chemotherapy," In : Chessin M. (Ed), *Antiviral Proteins in Higher Plants*, CRC Press, Boca Raton, FL, 1995, 161-174.
- [28] J.L. Mau, S.Y. Tsai, Y.H. Tseng, S.J. Huang "Antioxidant properties of hot water extracts from GA NODERMA T SUGAE Murrill. LWT," - *Food Science and Technology*, 2005, Volume 38, Issue 6, Pages 589-597
- [29] J. F. Clevenger, "Apparatus for volatile oil determination: description of New Type Clevenger," *Am Perf Ess Oil Review*, 1928, 467- 503.
- [30] Afnor, "Huiles essentielles. Échantillonnage et méthodes d'analyse (tome 1)," - *Monographies relatives aux huiles essentielles (tome 2. volumes 1 et 2) mars*, 2000.
- [31] H.L. Barnett, "Illustrated genera of imperfect fungi," 2nd ed., Minneapolis, Minn.: Burgess Publishing Co., 1930, p. 225.
- [32] C. M. A. Ellis, "The size at maturity and breeding season of sardines in southern Lake Tanganyika ," *Afr. J. Trop. Hydrobiol. Fish.*, 1971, 1(1):59-66.
- [33] P.E. Nelson, T.A. Toussou, W.F.O. Marasas, "*Fusarium* species-an illustrated manual for identification," *The pennsylvania State University press*, 1983, London.
- [34] K.H. Domsch, W. Gams, T.H. Anderson, "Compendium of soil fungi," London (UK), 1980, : Academic Press, 1th ed., p. 85
- [35] C. J. K. Wang, and R. A. Zabel, *Identification " Manual for Fungi from Utility Poles in the Eastern United States,"* Allen Press Inc, 1990, Lawrence, KS, U.S.A. 356 pp.
- [36] Y. A. Chabbert, G. L Daguét, " Techniques enbactériologie : antibiotiques en bactériologie médicale," Editeur: Flammarion, Médecine & Sciences, Sérologie bactérienne, 1985, p.244.
- [37] J.N.A. Eloff, " Sensitive and quick microplate method to determine the minimal inhibitory concentration of plant extracts for bacteria," *Planta Med.*, 64, 1998, 711-3.
- [38] K. Hostettmann, "Tout savoir sur le pouvoir des plantes, sources de médicaments ," 1997, Pierre Marcel Favre Editeur, Lausanne.
- [39] J.L. Salle, "Plantes médicinales et médecine traditionnelle d'Afrique," 1991, Karthala, Paris,
- [40] A. EL Ouali Lalami, F. El-Akhal, W. Ouedrhiri, F. Ouazzani Chahdi, R. Guemmouh, H. Greche, " Le thym vu au microscope – Composition chimique et activité antibactérienne des huiles essentielles de deux plantes aromatiques du centre nord marocain : *Thymus vulgaris* et *Thymus satureioides*," 2013, *La Gazette du laboratoire Maghreb*. N78, P 7. ; www.gazettelabo.ma
- [41] S. HMIRI, M. RAHOUTI, Z. HABIB, B. SATRANI, M. GHANMI et M. EL AJJOURI, "Evaluation du potentiel antifongique des huiles essentielles de *mentha pulegium* et d'*eucalyptus camaldulensis* dans la lutte biologique contre les champignons responsables de la détérioration des pommes en conservation," *Bulletin de la Société Royale des Sciences de Liège*, Vol. 80, 2011, p.824 - 836
- [42] E.L. Souza , T.L.M. Stamford, E.O. Lima, V.N. Trajano, "Effectiveness of ORIGANUM VULGARE L. essential oil to inhibit the growth of food spoiling yeasts," *Food Control*, Volume 18, 2007, Issue 5, Pages 409-413
- [43] M. Aberchane, M Fechtal, A Chaouch, T Bouayoune, "Influence de la durée et de la technique d'extraction sur le rendement et la qualité des huiles essentielles du cèdre de l'Atlas (*Cedrusatlantica manetti*)," *Annales de la recherche forestière au Maroc* ISSN 0483- 8009 CODEN AFRMA. : 2001, 34, 110-118.
- [44] S. Bouhdid, M. Idaomar, A. Zhiri, D. Baudoux, N. S Skali, J. Abrini, "*Thymus* essential oils: chemical composition and *in vitro* antioxidant and antibacterial activities," In: *Congrès International de Biochimie, 9-12 mai 2006, Agadir, Maroc*.

- [45] M. Bourkhiss, M. Hnach, T. Lakhlifi, A. Boughdad, A. Farah, B. Satrani, "Effet de l'Age et du Stade Végétatif sur la Teneur et la Composition Chimique des Huiles Essentielles de Thuya de Berbère," *Les technologies de laboratoire*, 2006, 6(23), 64-68.
- [46] F. Amarti, B. Satrani, M. Ghanmi, A. Farah, A. Aafi, L. Aarab, M. EL Ajjouri, et A. Chaouch, "Composition chimique et activité antimicrobienne des huiles essentielles de *Thymus algeriensis* Boiss. and Reut et *Thymus ciliatus* (Desf) Benth. du Maroc," *Biotechnol. Agron. Soc. Environ.*, 2010, 14 (1), 141-148.
- [47] G. Milhau, A. Valentin, F. Benoit, M. Mallié, J.M. Bastide, "In vitro antimalarial activity of eight essential oils," *J essent . oil Res*, 1997, 9 329 .
- [48] Mohammedi Z, Bachik S, Belkaroube N (2010). "Potentiel antifongique et anti aflatoxinogène des huiles essentielles d'une *Thymus fontanesii* Boiss and Reut," *Les technologies de laboratoire* : 2010, 5 (19), 10-15.
- [49] A. R. Khosravi, H. Shokri, S. Kermani, M. Dakhili, M. Madani, S. Parsa "Antifungal properties of *Artemisia sieberi* and *Origanum vulgare* essential oils against *Candida glabrata* isolates obtained from patients with vulvovaginal candidiasis," *Journal de Mycologie Médicale*, 2011, Volume 21, Issue 2, June 2011, Pages 93–99
- [50] Z. MOHAMMEDI, S. BACHIK, N. BELKAROUBE, "Potentiel antifongique et antiaflatoxinogène des huiles essentielles d'une plante endémique *Thymus fontanesii* Boiss. et Reut," *LA GAZETTE DU LABORATOIRE*, 2010, n°46 - octobre 2010. P8
- [51] M.K. Erhan, C. Bölükba, H. Ürü an "Biological activities of pennyroyal (*Mentha pulegium* L.) ," in broilers. *Livestock Science*, 2012, Volume 146, Issues 2–3, Pages 189–192.
- [52] Ghazghazi H, Chedia A, Weslati M, Trakhna F, Houssine S, Maaroufi A, Hasnaoui B "Chemical Composition and *in vitro* Antimicrobial Activities of *Mentha pulegium* Leaves Extracts against Foodborne Pathogens," *Journal of Food Safety*, 2013, Volume 33, Issue 3, pages 239–246.
- [53] D. Ouraini, A. Agoumi, M. Ismaili-Alaoui, K. Alaoui, Y. Cherrah, M. Amrani et M. Alaoui Belabbas, "Étude de l'activité des huiles essentielles de plantes aromatiques à propriétés antifongiques sur les différentes étapes du développement des dermatophytes," *Phytothérapie*, 2005, 4, 147-157.
- [54] D. Ouraini, A. Agoumi, M. Ismaili-Alaoui, K. Alaoui, Y. Cherrah, M.A. Alaoui et M.A. Belabbas, "Activité antifongique de l'acide oléique et des huiles essentielles de *Thymus saturejoides* L. et de *Mentha pulegium* L., comparée aux antifongiques dans les dermatoses mycosiques ," *Bulletin de la Société Royale des Sciences de Liège*, 2007, Vol. 80, 2011, p.824 – 836
- [55] B. Chebli, M. Achouri, L.M. Idrissi Hassani et M. Hmamouchi, "Chemical composition and antifungal activity of essential oils of seven Moroccan Labiatae against *Botrytis cinerea* Pers: Fr," *Journal of ethnopharmacology*, 2003. 89: 165–169.

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